



# Histone acetyltransferases are crucial regulators in NF- $\kappa$ B mediated inflammation

Massimo Ghizzoni<sup>1</sup>, Hidde J. Haisma<sup>1</sup>, Harm Maarsingh<sup>2</sup> and Frank J. Dekker<sup>1</sup>

<sup>1</sup> Department of Pharmaceutical Gene Modulation, Groningen Research Institute of Pharmacy, University of Groningen, Antonius Deusinglaan 1, 9713 AV Groningen, The Netherlands

<sup>2</sup> Department of Molecular Pharmacology, Groningen Research Institute of Pharmacy, University of Groningen, Antonius Deusinglaan 1, 9713 AV Groningen, The Netherlands

Post-translational modifications of proteins, such as acetylation, are important regulatory events in eukaryotic cells. Reversible acetylations of histones and non-histone proteins regulate gene expression and protein activity. Acetylation levels of proteins are regulated by a dynamic equilibrium between acetylation by (histone) acetyltransferases and deacetylation by (histone) deacetylases. Alterations in this equilibrium can result in pathological states. Inflammation is a physiological response that, under certain conditions, turns into a disease. This review focuses on the crucial regulatory roles of protein acetylation in NF- $\kappa$ B-mediated inflammation and the potential applications of small-molecule inhibitors of acetylation for the treatment of inflammatory diseases.

All cellular processes are regulated by a complex and dynamic network of signals and interactions that allows cells to respond to alteration in the cellular environment. This network is, among others, regulated by post-translational modifications such as acetylation, methylation phosphorylation and ubiquitination. These modifications alter protein–protein interactions, modulate enzyme activity and function as regulatory switches for many cellular processes. Histones are small proteins around which DNA is wrapped to form the basic packaging unit of DNA called the ‘nucleosome’. Post-translational modifications of histones modify their interactions with DNA and with other proteins, and change the chromatin structure to expose promoter regions for binding of transcription factors.

Inflammation is a physiological response, triggered by physical, biological or chemical stimuli, which is necessary for cell survival. Under certain circumstances, however, the inflammatory response is inappropriate or excessive, which leads to pathological states. Abnormal and chronic inflammatory responses have been associated with various diseases including asthma, cancer, diabetes and neurodegenerative diseases.

It has been demonstrated clearly that post-translational modifications of proteins play a crucial part in regulating the intensity, the duration and the specificity of inflammatory responses. This review focuses on the role of histone and non-histone protein acetylations by histone acetyltransferases (HATs) in nuclear factor  $\kappa$ B (NF- $\kappa$ B)-mediated inflammation. Furthermore, we discuss the inhibition of acetylation by small-molecule HAT inhibitors as a potential therapeutic approach to control inflammation.

## Protein acetylation

Reversible acetylation of histones and other proteins is modulated by the activity of HATs and histone deacetylases (HDACs). These enzymes catalyze, respectively, the introduction or removal of acetyl groups from  $\epsilon$ -amino functionalities of specific lysine residues. HATs are classified based on their cellular localization in nuclear HATs (type A) and cytoplasmatic HATs (type B) [1]. Several nuclear HATs have been identified, whereas only one cytoplasmatic HAT (HAT 1) has been described so far. HATs have been divided into five families based on their primary structure homology. The three families that have been studied extensively are: the GNAT (GCN5-related N-acetyltransferase) family, represented by GCN5 (general control nonderepressible 5) and PCAF (p300/CBP associated factor); the p300/CBP family, including p300 and CBP

Corresponding author: Dekker, F.J. (f.j.dekker@rug.nl)

(CREB-binding protein); and the MYST family, which includes Tip60 (TAT-interacting protein 60). The acronym MYST derives from the four founding members of this HAT family: mammalian MOZ, yeast Y bf2/Sas3 and Sas2, and mammalian Tip60.

### Role of acetylation in NF- $\kappa$ B-mediated inflammation

The NF- $\kappa$ B transcription factors encompass a family of inducible transcription factors that play a crucial part in the expression of numerous genes that are involved in immune and inflammatory responses and in cell survival [2]. NF- $\kappa$ B transcription factors exist in homo- or hetero-dimeric complexes consisting of different members of the Rel family of proteins. The most prevalent and best studied of these complexes is the p50–p65 heterodimer. In resting cells, p50–p65 is present in the cytoplasm in an inactive form, bound to inhibitory proteins known as I $\kappa$ Bs (Fig. 1). Upon stimulation by specific inducers such as inflammatory cytokines [tumor necrosis factor (TNF) and interleukin-1 (IL-1)], bacterial products (lipopolysaccharide) or oxidative stress (H<sub>2</sub>O<sub>2</sub>), the I $\kappa$ Bs are phosphorylated, ubiquitinated and degraded. Degradation of I $\kappa$ Bs results in the release of the p50–p65 dimer, which translocates into the nucleus, followed by specific upregulation of gene expression. Owing to the central role of NF- $\kappa$ B in inflammation, the NF- $\kappa$ B pathway has been recognized as a target for therapeutic intervention.

#### Acetylation of NF- $\kappa$ B complex

Several regulatory mechanisms control the NF- $\kappa$ B response to specific stimuli. Among these, post-translational modifications play an important role and regulate different functions of NF- $\kappa$ B. Post-translational modifications mutually influence each other, which results in a complex pattern of modifications that all determine the output and the duration of responses upon specific stimulations of NF- $\kappa$ B. In this review we focus on reversible acetylation of NF- $\kappa$ B and histones connected to its target genes. Recently, it has been described that acetylations of specific lysine residues of NF- $\kappa$ B subunits play distinct roles in the regulation of its transcriptional capacity, its DNA-binding ability and duration of its actions (Fig. 1).

It has been shown that the p65 NF- $\kappa$ B subunit (also known as RelA) is acetylated at specific sites by different HATs. Chen *et al.* reported acetylation at lysine residues 218, 221 and 310 by the HATs p300 and CBP [3]. Acetylation at Lys 221 increases the binding affinity of the NF- $\kappa$ B complex to the DNA  $\kappa$ B enhancer. Furthermore, acetylation at Lys 221, alone or in combination with Lys 218, impairs assembly of NF- $\kappa$ B with newly synthesized I $\kappa$ B $\alpha$ , which extends the duration of the NF- $\kappa$ B activity. By contrast, deacetylation by HDAC3 stimulates I $\kappa$ B $\alpha$  binding, which promotes NF- $\kappa$ B export from the nucleus to the cytoplasm [4]. Acetylation at Lys 310 is required for full transcriptional activity of p65, but does not affect the DNA binding or its assembly with I $\kappa$ B $\alpha$ . In a subsequent study, it was shown that acetylation on this position provides a binding site for binding of the bromodomain of the transcriptional coactivator Brd4 [5]. Recent work by Yang *et al.* showed that acetylation at Lys 310 enhances the transcriptional activity of p65 by impairing the methylation of lysine residues 314 and 315, which is important for the ubiquitination and degradation of chromatin-associated p65 [6]. The HDAC Sirtuin 1 (SIRT1) deacetylates p65 at Lys 310 and thus terminates NF- $\kappa$ B-dependent gene expression [7]. Thus, acetylations of NF- $\kappa$ B on residues 218, 221 and 310 increase its transcriptional activity.

By contrast, acetylation at other positions decreases the NF- $\kappa$ B transcriptional activity. Acetylations on the lysine residues 122 and 123 by p300 and PCAF reduce binding of NF- $\kappa$ B to the DNA  $\kappa$ B enhancer and facilitate binding to I $\kappa$ B $\alpha$  and subsequent export from the cytoplasm [8]. Other acetylations have no direct effect on transcriptional activity. Acetylation at lysine residues 314 and 315 by p300 do not affect the general transcriptional activity of the NF- $\kappa$ B complex [9,10]. Nevertheless, these acetylations modulate the expression of specific sets of genes, which indicate that site-specific acetylations of p65 regulate the specificity of the NF- $\kappa$ B-dependent gene expression.

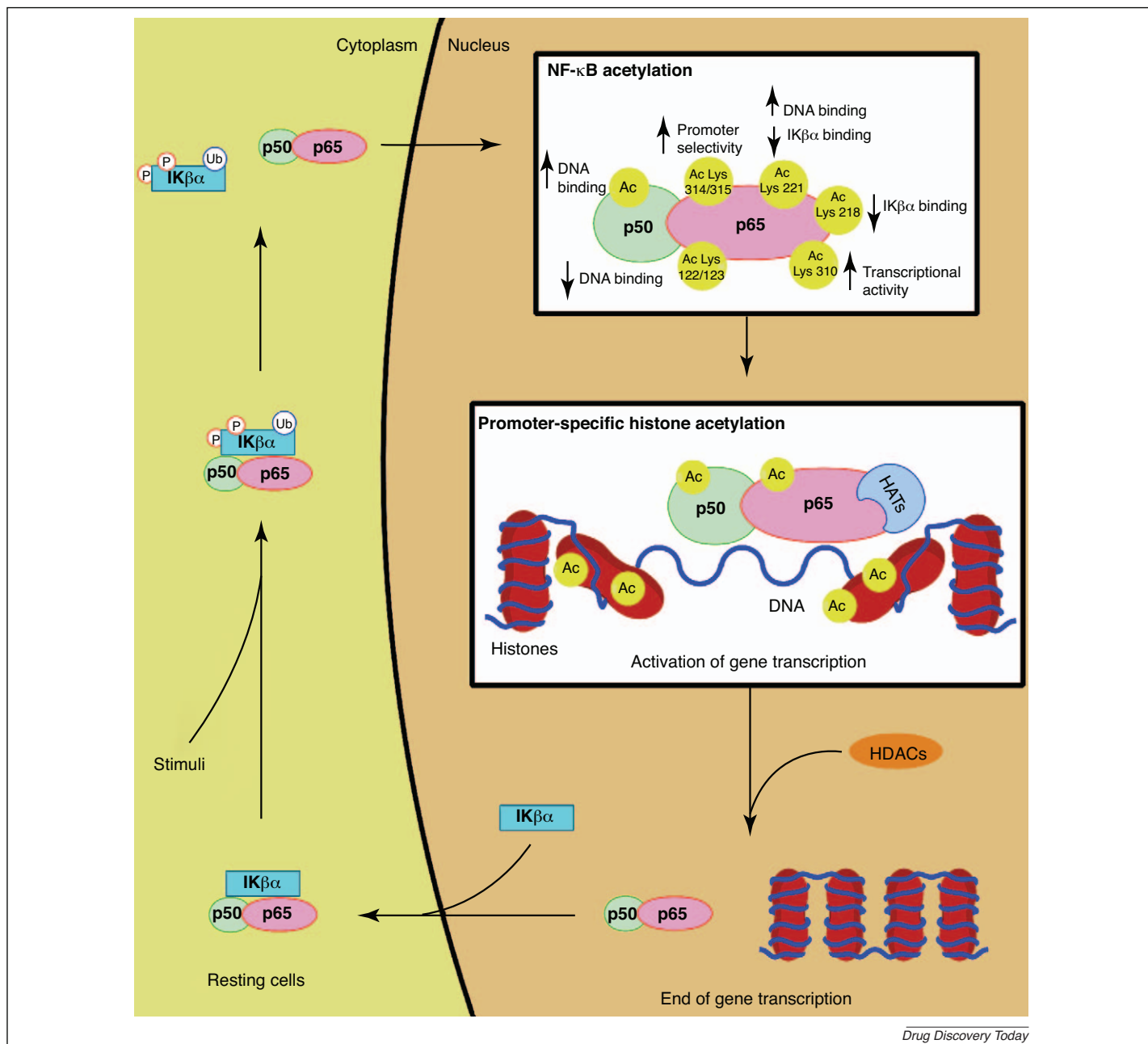
The p50 subunit of NF- $\kappa$ B is also subject to stimulus-induced acetylation. *In vitro*, p50 is acetylated at lysine residues 431, 440 and 441 [11]. It has been shown that acetylation by p300 increases binding of p50 to the DNA and enhances the transcriptional activation by NF- $\kappa$ B [12].

Further evidence for the importance of direct acetylations of NF- $\kappa$ B subunits as regulatory mechanisms is the observation that diverse cofactors can regulate NF- $\kappa$ B transactivation by modulating p50–p65 acetylation levels. For example, the transcriptional repressor Daxx impairs the transcriptional activity of NF- $\kappa$ B by binding to p65 and interferes with its acetylation [13]. By contrast, the transcription coactivator Stat3 maintains constitutive NF- $\kappa$ B activity in inflammation-induced cancers by promoting p300-mediated acetylation of the p65 subunit [14].

Taking these data together it is concluded that direct acetylation and deacetylation of specific lysine residues in the p50 and p65 subunits of NF- $\kappa$ B play a crucial part in the regulation of different NF- $\kappa$ B functions. Acetylations on different sites have different effects on NF- $\kappa$ B transcriptional activity and NF- $\kappa$ B-dependent gene expression. Several studies show that inhibition of deacetylation extends NF- $\kappa$ B transcriptional activity in response to specific stimuli, whereas other studies demonstrate that inhibition of deacetylation decreases NF- $\kappa$ B transcriptional activity [2,15,16]. This indicates that the resulting effect of both HAT and HDAC inhibition depends on the selectivity for specific NF- $\kappa$ B acetylation sites. The crucial role of NF- $\kappa$ B acetylation and deacetylation in the regulation of NF- $\kappa$ B-mediated gene expression raises the idea to modulate inflammatory responses by modulating NF- $\kappa$ B acetylation levels with HAT and HDAC inhibitors. It has been shown that NF- $\kappa$ B acetylations are mediated by the HATs such as p300, CBP and PCAF, which indicates that small-molecule inhibitors of these HATs will modulate NF- $\kappa$ B signaling via direct inhibition of NF- $\kappa$ B acetylation as well as via modulation of histone acetylation.

#### Acetylation of co-activators of NF- $\kappa$ B

Several cofactors that are involved in the regulation of the NF- $\kappa$ B transactivation are subject to direct acetylations by HATs. These acetylations modulate their interactions with the NF- $\kappa$ B subunits. Such acetylations have been described for poly(ADP-ribose) polymerase-1 (PARP-1), Signal Transducers and Activators of Transcription 1 (Stat1) and the glucocorticoid receptors. PARP-1 is a coactivator of NF- $\kappa$ B that has been demonstrated to have a role in inflammatory disorders. PARP-1 is acetylated upon inflammatory stimulation on specific lysine residues by p300 and CBP and deacetylated by HDACs 1–3 [17]. Acetylation of PARP-1 is required for the interaction with p50, which results in coactivation of NF- $\kappa$ B in response to inflammatory stimuli. Stat1 is a transcription factor

**FIGURE 1**

Importance of acetylation in NF-κB-mediated inflammation. In resting cells, p50-p65 subunits are present in the cytoplasm in an inactive form, bound to the inhibitory protein IκBα. Upon stimulation IκBα is phosphorylated, ubiquitinated and degraded. The p50-p65 dimer is released and translocates into the nucleus where both subunits are acetylated in different positions. Acetylations of p50-p65 regulate its transcriptional capacity, its DNA-binding ability and the duration of its action. p50-p65 subunits recruit HATs to the target gene. Gene-specific histone acetylations by HATs change the chromatin structure exposing promoter regions for binding of p50-p65.

that can modulate NF-κB activity. Acetylation of Stat1 by HATs, such as CBP, results in binding to the p65 subunit of NF-κB and consequently the reduction of NF-κB signaling [18]. The action of glucocorticoids involves acetylation of the glucocorticoid receptor. The deacetylated form of the glucocorticoid-bound glucocorticoid receptor can suppress NF-κB activity [19].

#### Acetylation of histones in NF-κB-mediated gene transcription

Dynamic acetylations and deacetylations play an important role in the regulation of gene transcription resulting from activation of

the NF-κB pathway. Upon DNA binding the NF-κB subunits can recruit HATs and HDACs to the target gene promoter to change the acetylation profiles of the histones. It has been shown that the transcriptional activation domain of p65 interacts with the N-terminal and C-terminal domain of the HATs CBP and p300 [20]. These HATs function as co-activators for gene transcription. It has been demonstrated that the presence of HATs, such as CBP, p300 and PCAF, at the gene promoter is essential for NF-κB-mediated gene expression [21]. By contrast, it has also been shown that p65 interacts with HDACs, such as HDAC-1 and SIRT6, that function as

TABLE 1

**Promoter-specific hyperacetylation of NF- $\kappa$ B-dependent genes**

Gene	Stimulus	Cell type	Acetylation site	HAT involved	Ref.
GM-CSF	IL-1 $\beta$	A549	Histone H4 Lys 8, 12	Unknown	[24]
Eotaxin-1	TNF- $\alpha$	HASMC	Histone H4 Lys 5, 12	PCAF	[26]
Interleukin-8	Legionella	Human lung epithelial	Histone H3 Lys 14, Histone H4	p300/CBP	[31]
	Listeria	HUVEC	Histone H3 Lys 14, Histone H4 Lys 8	CBP	[32]
	LPS	HUT-78 and Jurkat and monocytes	Histone H4	Unknown	[33]
	H <sub>2</sub> O <sub>2</sub> and PM <sub>10</sub>	A549	Histone H4	Unknown	[34]
Interleukin-6	TNF- $\alpha$	L929sA HEK293T	Histone acetylation	p300/CBP	[27]
ELAM	TNF- $\alpha$	L929sA HEK293T	Histone acetylation	p300/CBP	[27]
CXCL10	TNF- $\alpha$ and INF- $\gamma$	ASM	Histone H4	CBP	[28]
E-selectin	TNF- $\alpha$	HUVECs	Histone H3 Lys 9, 14, Histone H4 Lys 5, 8, 12	PCAF p300	[29]
Cox-2	Bradykinin	ASM	Histone H4 Lys 5, 8, 16	Unknown	[30]
	IL-1 $\beta$	ASM	Histone H4 Lys 8	Unknown	[30]

repressors of gene transcription by deacetylating histones at promoters of NF- $\kappa$ B target genes [22,23].

The importance of promoter-specific histone acetylation by HATs has been demonstrated for many NF- $\kappa$ B target genes (Table 1). HATs are involved in gene transcription of granulocyte-macrophage colony-stimulating factor (GM-CSF), which is a protein that is involved in physiological and pathological inflammatory processes. IL-1 $\beta$  stimulates GM-CSF production through activation of the NF- $\kappa$ B pathway. The GM-CSF production is associated with increased acetylation at Lys 8 and Lys 12 of histone H4 connected to the GM-CSF promoter [24]. Another example is CCL11 (also known as eotaxin-1), which is an eosinophil chemoattractant with relevance in allergic diseases such as asthma. CCL11 transcription is increased upon stimulation with TNF- $\alpha$ . NF- $\kappa$ B is the key transcription factor for CCL11. In the TNF- $\alpha$ -induced CCL11 transcription, PCAF is recruited, upon phosphorylation, to the CCL11 promoter. PCAF increases histone H4 acetylation at Lys 5 and Lys 12, which promotes p65 association with DNA and the transcription of the CCL11 gene. Histone H4 acetylation is markedly reduced after treatment with  $\beta_2$ -agonists and glucocorticoids. This suggests that one of the mechanisms of actions of these medicines is the indirect decrease of histone acetylation, which inhibits inflammatory gene transcription [25,26].

IL-6 is another important mediator of inflammatory responses. Overexpression of CBP and p300 potentiates basal and TNF- $\alpha$ -induced IL-6 promoter activation via interactions with the NF- $\kappa$ B subunit p65. A similar mechanism is proposed for the endothelial leukocyte adhesion molecule (ELAM), which is another NF- $\kappa$ B-dependent promoter [27]. Interferon- $\gamma$ -inducible protein-10 (CXCL10) is a chemokine implicated in the pathophysiology of asthma and COPD (chronic obstructive pulmonary disease). Stimulation of asthmatic airway smooth muscle cells with TNF- $\alpha$  or interferon (IFN)- $\gamma$  induces recruitment of CBP and acetylation of histone H4, but not H3 at the CXCL10 promoter, which consequently increases its transcriptional activity [28]. Another gene that is expressed upon inflammatory stimulation is E-selectin. E-selectin is an adhesion molecule that is rapidly expressed by endothelial cells activated upon inflammatory stimulation. Induc-

tion of E-selectin gene expression by TNF- $\alpha$  is associated with hyperacetylation of histones H3 and H4 connected to the E-selectin promoter by the HATs PCAF and p300 [29]. Another gene where expression is induced in response to inflammation is cyclooxygenase 2 (COX-2). Transactivation of COX-2 gene expression involves selective hyperacetylation of histone H4 connected to the COX-2 promoter in a stimulus-specific manner. Stimulation with bradykinin causes acetylation at Lys 5, 8 and 16, whereas stimulation with IL-1 $\beta$  causes only acetylation at Lys 8 [30]. This demonstrates that specific stimuli induce a distinct histone acetylation pattern, which regulates distinct gene transcription patterns in response to specific inflammatory stimuli.

#### Histone acetylation in IL-8 gene expression

Histone acetylation has an important role in the activation of IL-8 gene expression in immune responses triggered by bacterial infections. For example, IL-8 expression in *Legionella pneumophila* infected lung cells involves increased recruitment of p300 and CBP, as well as enhanced acetylation of histones H3 and H4 at the IL-8 gene promoter [31]. The same is observed upon infection by intracellular *Listeria monocytogenes*, which also induces acetylation of histone H3 and H4 at the IL-8 promoter, consequently increasing IL-8 expression. By contrast, *Listeria*-induced IFN- $\gamma$  gene expression does not require changes in the histone acetylation at the IFN- $\gamma$  promoter. Furthermore, the HDAC inhibitor trichostatin A increases *Listeria*-induced expression of IL-8, but not of IFN- $\gamma$ -induced IL-8 expression. This demonstrates that histone acetylation regulates the gene expression patterns in response to specific stimuli [32]. Another stimulus that increases IL-8 expression is lipopolysaccharides (LPS). LPS-induced IL-8 gene transcription is associated with acetylation of histone H4 at the IL-8 promoter. Interestingly, glucocorticoids suppress IL-8 expression, which is accompanied by the reduction of histone acetylation of the IL-8 promoter [33]. By contrast, IL-8 expression and release is also increased upon oxidative stress induced by H<sub>2</sub>O<sub>2</sub> and environmental particulate matter (PM<sub>10</sub>). It has been shown that H<sub>2</sub>O<sub>2</sub> and PM<sub>10</sub> augment the intrinsic HAT activity in alveolar epithelial cells leading to increased histone H4 acetylation at the IL-8 gene

promoter [34]. These findings demonstrate that IL-8 expression is positively correlated with histone acetylation.

## Role of acetylation in disease

### Asthma and COPD

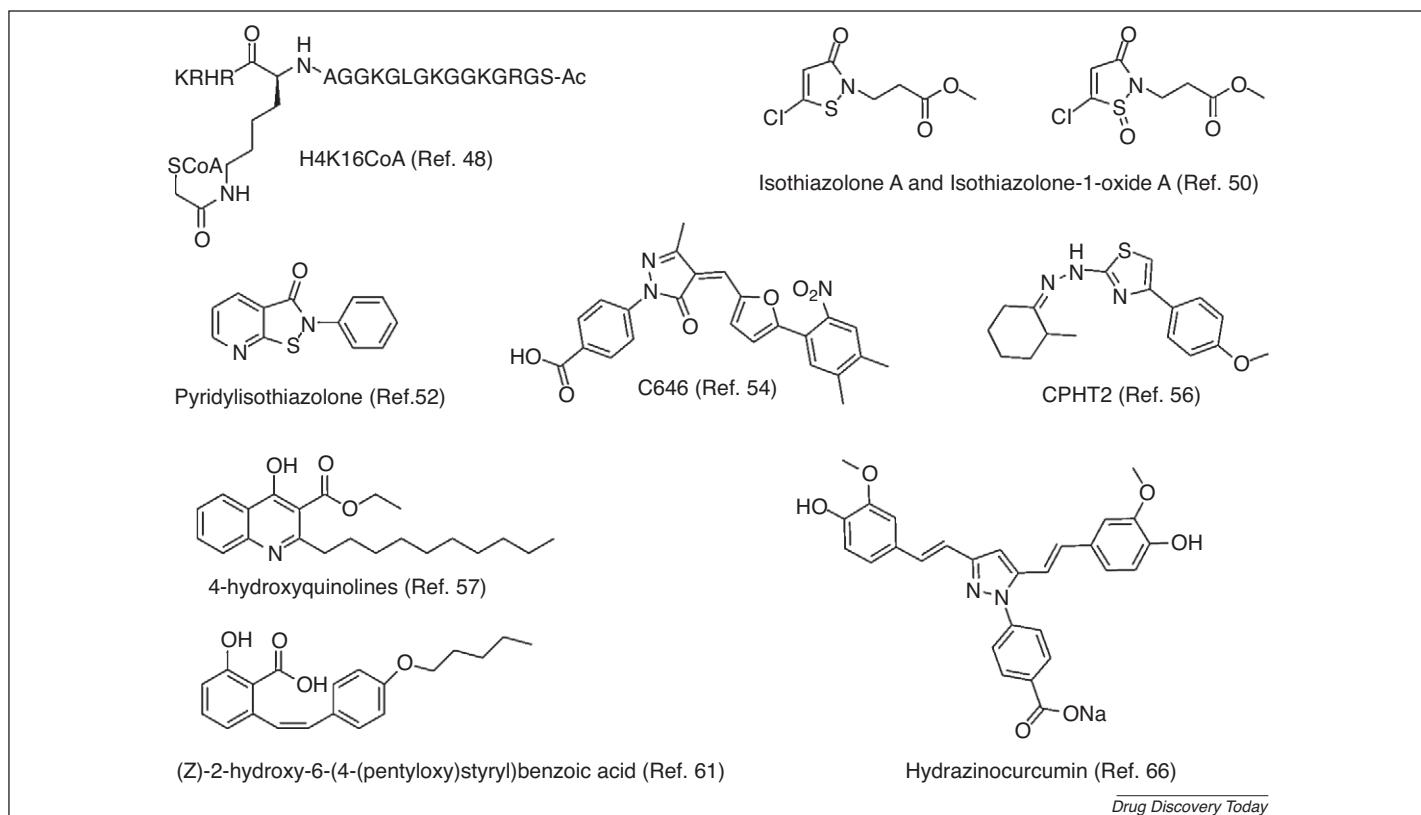
Inflammatory lung diseases such as asthma and COPD are associated with the expression of multiple inflammatory genes in the lungs. Increasing evidence demonstrates that the balance between HAT and HDAC activity is altered in these diseases. A recent study showed significant reduction of HDAC activity and increased HAT activity in peripheral blood mononuclear cells from children with allergic asthma. The intensity of these alterations was positively correlated to bronchial hyper-responsiveness [35]. In another study, it has been shown that bronchial biopsies from subjects with asthma contain reduced HDAC1 and HDAC2 expression and decreased HDAC activity. By contrast, HAT activity was increased, although no difference in expression of HATs (PCAF and CBP) was found [36]. Furthermore, it has been found that HAT activity is increased and HDAC activity is decreased in alveolar macrophages of subjects with asthma compared with healthy subjects [37]. The HAT activity was reduced to normal levels in patients that were treated with glucocorticoids, which indicates that glucocorticoids regulate inflammatory responses indirectly by the reduction of HAT activity.

The inflammatory lung disease COPD is also characterized by a disturbed HAT/HDAC balance. A study with lung tissue samples of COPD patients showed reduced HDAC activity and expression, and increased histone H4 acetylation at the IL-8 promoter. In

contrast to asthma, HAT activity in COPD patients was not increased in comparison with healthy subjects [38]. The reduced HDAC expression and activity measured in COPD are also responsible for the steroid resistance associated with this disease [19]. Taken together, these findings indicate that a shift of the HAT/HDAC balance towards HAT activity could underlie the increased expression of inflammatory genes in inflammatory lung diseases, which is a mechanism that has been described in several review articles [39–41]. This demonstrates that inhibition of HAT activity has potential for the treatment of such inflammatory diseases.

### Diabetes

Inflammatory processes have a role in the pathogenesis of diabetes. This has been shown in experiments with high glucose conditions that mimic diabetes. These experiments demonstrated activation of NF- $\kappa$ B-dependent gene transcription of inflammatory genes in monocytes *in vitro*. This activation proceeds through recruitment of p65 and HATs to the TNF- $\alpha$  and COX-2 promoters, with concomitant increases in histone H3 and H4 acetylation. Furthermore, increased histone H3 acetylation at TNF- $\alpha$  and COX-2 promoters has been found in human blood monocytes from type 1 and type 2 diabetic subjects in comparison with non-diabetic subjects [42]. A study in endothelial cells and vascular tissue demonstrated that glucose causes upregulation of p300 accompanied by increased histone acetylation and expression of extracellular matrix proteins and vasoactive factors. These proteins are responsible for alterations of endothelial and vascular structure and function in organs, which are affected by chronic diabetic



Drug Discovery Today

FIGURE 2

Recent small molecule inhibitors of histone acetyltransferases.



complications [43]. These results suggest that inhibition of histone acetylation might be a valuable strategy to suppress inflammatory responses in diabetes.

### Neurodegenerative diseases

Neurodegenerative diseases, such as Alzheimer's and Parkinson's disease and amyotrophic lateral sclerosis, are characterized by slow and progressive dysfunction and loss of neurons in the central nervous system. Several findings support the theory that inflammation contributes to the development of the neurodegeneration by activating chronic immune responses, in particular by microglia and astroglia. Highly activated glial cells express pro-inflammatory cytokines and chemokines that increase excitotoxicity on neurons and are responsible for neuronal apoptosis [44,45]. The inflammatory response of glial cells involves the activation of different signaling pathways, including NF- $\kappa$ B, which is accompanied by histone hyperacetylation [46–48]. There are many examples in the literature of small molecules that provide protection against glial-cell-induced neurodegeneration by blocking the NF- $\kappa$ B pathway [49,50]. These findings raise the idea that HAT inhibitors could also find a therapeutic application in this field.

### HAT inhibitors in inflammation

Small-molecule HAT inhibitors that are subtype-selective and cell-permeable are essential tools to evaluate the role of HATs in inflammation. Several classes of HAT inhibitors have been reviewed previously [51,52]. The most potent and selective HAT inhibitors are the so-called bisubstrate inhibitors, which include the histone peptide and CoA (Fig. 2). These inhibitors are remarkably selective and potent but their lack of cell permeability limits their applicability [53,54]. High-throughput screening led to the identification of isothiazolones as potent HAT inhibitors [55,56]. Unfortunately, the high reactivity has limited their applications in cell-based studies [57,58]. Nevertheless, this class of covalent inhibitors provides opportunities for activity-based protein profiling of HATs in cell lysates [59]. Recently, a promising HAT inhibitor was discovered using virtual ligand screening [60]. This compound is a potent and selective inhibitor of p300 and it can reduce histone acetylation and cancer cell growth. Another group used phenotypic screening for the identification of new HAT inhibitors [61–63]. Nevertheless, none of the described compounds has been used to investigate the effect of small-molecule inhibition of acetylation on inflammation.

The screening of natural products has led to the identification of several HAT inhibitors (Fig. 3). Interestingly, these compounds originate from plants known in traditional medicine to have anti-inflammatory effects. For example, anacardic acid (AA) has been described as an inhibitor of the HATs p300/CBP and PCAF [64]. Interestingly, AA has anti-inflammatory properties. Sung *et al.* reported that AA inhibits acetylation of p65, and suppresses inducible and constitutive NF- $\kappa$ B activation with consequent reduction of NF- $\kappa$ B-dependent gene expression. The downregulation of p300 by siRNA abrogated the effects of AA on the NF- $\kappa$ B pathway, which suggests that the HAT inhibitory effect is essential for the anti-inflammatory properties of this compound [65]. Because of its promising characteristics, AA has been used to design new HAT inhibitors [66–68].

Another natural product that has recently been described to inhibit the HAT p300 [69] is plumbagin. Plumbagin also suppresses NF- $\kappa$ B activation induced by different inflammatory stimuli and downregulates NF- $\kappa$ B-dependent gene expression [70]. Interestingly, the intensively investigated natural product curcumin is an inhibitor of p300/CBP [71]. Its synthetic derivative, hydrazinocurcumin, also inhibits HATs [72]. Curcumin reduces NF- $\kappa$ B activation induced by a variety of stimuli and downregulates NF- $\kappa$ B-dependent expression of many inflammatory genes via suppression of TNF- $\alpha$ -induced p65 acetylation [73]. Furthermore, the natural products gallic acid and epigallocatechin-3-gallate also possess anti-inflammatory properties. Recently, these compounds were described as non-selective inhibitors of HATs [74,75]. They suppress p65 acetylation and abrogate NF- $\kappa$ B activation in response to different inflammatory stimuli. It should, however, be noted that epigallocatechin-3-gallate also inhibits phosphorylation of several proteins such as MAP kinases [76]. Finally, the natural product garcinol is a potent inhibitor of p300 and PCAF and also possesses anti-inflammatory properties [77]. It has been shown that garcinol inhibits constitutive and induced NF- $\kappa$ B activity and downregulates NF- $\kappa$ B-dependent genes [78]. Microarray analysis showed that garcinol inhibited the expression of many disease-related genes [79]. Taking these data together it is tempting to speculate that the anti-inflammatory and HAT inhibitory properties of these natural products are directly connected. Nevertheless, it should be noted that many HAT inhibitors also influence other protein targets that could cause their activity. In conclusion, the development of selective inhibitors for HAT (iso)-enzymes that specifically downregulate expression of distinct genes remains a major challenge.

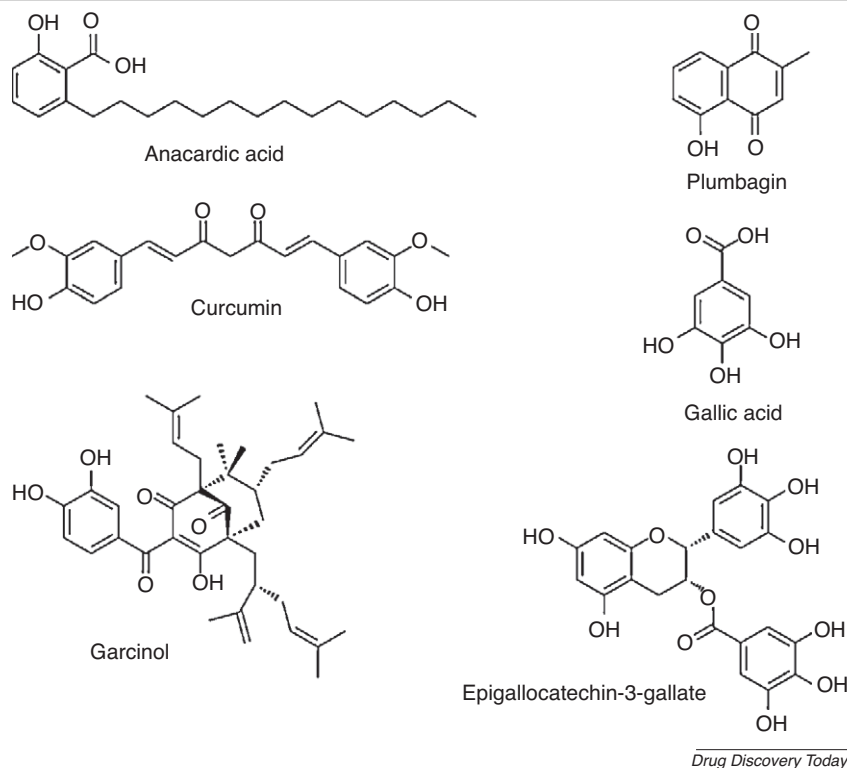
### Conclusions

#### *Acetylations and deacetylations are crucial regulators in inflammation*

Recent studies clearly demonstrate that inflammation requires acetylation of histone and non-histone proteins. The crucial regulatory role of acetylation has been extensively demonstrated for the NF- $\kappa$ B pathway. Acetylation regulates the function of many proteins that are involved in the NF- $\kappa$ B pathway. Direct acetylation and deacetylation of the NF- $\kappa$ B subunits p50 and p65 regulate their signaling output. Furthermore, it has been demonstrated that acetylation of co-activators of the NF- $\kappa$ B pathway is required for transcriptional activation. Finally, it has been shown that acetylation of the histones that are connected to NF- $\kappa$ B target genes exposes promoter regions for NF- $\kappa$ B binding. This allows transcriptional activation of pro-inflammatory genes. It should, however, be noted that the regulatory machinery of inflammation is incredibly complex and many questions remain to be answered. It is, for example, not always clear which HATs are required for which acetylations in the NF- $\kappa$ B pathway. Moreover, the relative importance of the different acetylations by different HATs for activation of the NF- $\kappa$ B remains to be investigated.

#### *Small-molecule inhibitors of HATs as tools to suppress inflammation*

Inappropriate or excessive inflammatory responses that underlie many diseases are often accompanied by hyperacetylation of proteins that are involved in the NF- $\kappa$ B pathway. HATs are crucial

**FIGURE 3**

Natural product inhibitors of histone acetyltransferases.

for acetylation of many proteins that are involved in the NF- $\kappa$ B pathway. The commonly observed hyperacetylation in inflammation demonstrates that small-molecule inhibitors of HATs have potential to suppress inflammation. In addition, HAT inhibitors are valuable tools in pharmacological studies on inflammation. The number of HAT inhibitors has been constantly growing in recent years; however, their selectivity and potency is still limited.

The development of novel HAT inhibitors with improved potency and selectivity is therefore urgently required to develop these compounds as anti-inflammatory drugs.

### Acknowledgments

We thank the COST action Epigenetics: from bench to bedside (TD0905) for financial support.

### References

- Roth, S.Y. *et al.* (2001) Histone acetyltransferases. *Annu. Rev. Biochem.* 70, 81–120
- Baldwin, A.S., Jr (1996) The NF- $\kappa$ B and I  $\kappa$ B proteins: new discoveries and insights. *Annu. Rev. Immunol.* 14, 649–683
- Chen, L.F. *et al.* (2002) Acetylation of RelA at discrete sites regulates distinct nuclear functions of NF- $\kappa$ B. *EMBO J.* 21, 6539–6548
- Chen, L. *et al.* (2001) Duration of nuclear NF- $\kappa$ B action regulated by reversible acetylation. *Science* 293, 1653–1657
- Huang, B. *et al.* (2009) Brd4 coactivates transcriptional activation of NF- $\kappa$ B via specific binding to acetylated RelA. *Mol. Cell. Biol.* 29, 1375–1387
- Yang, X.D. *et al.* (2010) Functional interplay between acetylation and methylation of the RelA subunit of NF- $\kappa$ B. *Mol. Cell. Biol.* 30, 2170–2180
- Yeung, F. *et al.* (2004) Modulation of NF- $\kappa$ B-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J.* 23, 2369–2380
- Kiernan, R. *et al.* (2003) Post-activation turn-off of NF- $\kappa$ B-dependent transcription is regulated by acetylation of p65. *J. Biol. Chem.* 278, 2758–2766
- Buerki, C. (2008) Functional relevance of novel p300-mediated lysine 314 and 315 acetylation of RelA/p65. *Nucleic Acids Res.* 36, 1665–1680
- Rothgier, K.M. *et al.* (2010) Acetylation of p65 at lysine 314 is important for late NF- $\kappa$ B-dependent gene expression. *BMC Genomics* 11, 22–32
- Furia, B. *et al.* (2002) Enhancement of nuclear factor- $\kappa$ B acetylation by coactivator p300 and HIV-1 Tat proteins. *J. Biol. Chem.* 277, 4973–4980
- Deng, W.G. and Wu, K.K. (2003) Regulation of inducible nitric oxide synthase expression by p300 and p50 acetylation. *J. Immunol.* 171, 6581–6588
- Park, J. *et al.* (2007) Inhibition of NF- $\kappa$ B acetylation and its transcriptional activity by Daxx. *J. Mol. Biol.* 368, 388–397
- Lee, H. *et al.* (2009) Persistently activated Stat3 maintains constitutive NF- $\kappa$ B activity in tumors. *Cancer Cell.* 15, 283–293
- Leoni, F. *et al.* (2005) The histone deacetylase inhibitor ITF2357 reduces production of pro-inflammatory cytokines *in vitro* and systemic inflammation *in vivo*. *Mol. Med.* 11, 1–12
- Faraco, G. *et al.* (2009) Histone deacetylase (HDAC) inhibitors reduce the glial inflammatory response *in vitro* and *in vivo*. *Neurobiol. Diseases* 36, 269–279
- Hassa, P.O. *et al.* (2005) Acetylation of poly(ADP-ribose) polymerase-1 by p300/CREB-binding protein regulates coactivation of NF- $\kappa$ B-dependent transcription. *J. Biol. Chem.* 280, 40450–40464
- Kramer, O.H. *et al.* (2006) Acetylation of Stat1 modulates NF- $\kappa$ B activity. *Genes Dev.* 20, 473–485
- Ito, K. *et al.* (2006) Histone deacetylase 2-mediated deacetylation of the glucocorticoid receptor enables NF- $\kappa$ B suppression. *J. Exp. Med.* 203, 7–13
- Gerritsen, M.E. *et al.* (1997) CREB-binding protein/p300 are transcriptional coactivators of p65. *Proc. Natl. Acad. Sci. U. S. A.* 94, 2927–2932
- Sheppard, K.A. *et al.* (1999) Transcriptional activation by NF- $\kappa$ B requires multiple coactivators. *Mol. Cell. Biol.* 19, 6367–6378
- Ashburner, B.P. *et al.* (2001) The p65 (RelA) subunit of NF- $\kappa$ B interacts with the histone deacetylase (HDAC) corepressors HDAC1 and HDAC2 to negatively regulate gene expression. *Mol. Cell. Biol.* 21, 7065–7077

- 23 Kawahara, T.L. *et al.* (2009) SIRT6 links histone H3 lysine 9 deacetylation to NF-kappaB-dependent gene expression and organismal life span. *Cell* 136, 62–74
- 24 Ito, K. *et al.* (2000) Glucocorticoid receptor recruitment of histone deacetylase 2 inhibits interleukin-1beta-induced histone H4 acetylation on lysines 8 and 12. *Mol. Cell Biol.* 20, 6891–6903
- 25 Clarke, D.L. *et al.* (2008) PKCbetaII augments NF-kappaB-dependent transcription at the CCL11 promoter via p300/CBP-associated factor recruitment and histone H4 acetylation. *J. Immunol.* 181, 3503–3514
- 26 Nie, M. *et al.* (2005) beta2-Adrenoceptor agonists, like glucocorticoids, repress eotaxin gene transcription by selective inhibition of histone H4 acetylation. *J. Immunol.* 175, 478–486
- 27 Vanden Berghe, B.W. *et al.* (1999) The nuclear factor-kappaB engages CBP/p300 and histone acetyltransferase activity for transcriptional activation of the interleukin-6 gene promoter. *J. Biol. Chem.* 274, 32091–32098
- 28 Clarke, D.L. *et al.* (2010) TNFalpha and IFNgamma synergistically enhance transcriptional activation of CXCL10 in human airway smooth muscle cells via STAT-1, NF-kappaB, and the transcriptional coactivator CREB-binding protein. *J. Biol. Chem.* 285, 29101–29110
- 29 Edelstein, L.C. *et al.* (2005) Chromatin modification and the endothelial-specific activation of the E-selectin gene. *J. Biol. Chem.* 280, 11192–11202
- 30 Nie, M. *et al.* (2003) Transcriptional regulation of cyclooxygenase 2 by bradykinin and interleukin-1beta in human airway smooth muscle cells: involvement of different promoter elements, transcription factors, and histone h4 acetylation. *Mol. Cell Biol.* 23, 9233–9244
- 31 Schmeck, B. *et al.* (2008) Histone acetylation and flagellin are essential for *Legionella pneumophila*-induced cytokine expression. *J. Immunol.* 181, 940–947
- 32 Schmeck, B. *et al.* (2005) Intracellular bacteria differentially regulated endothelial cytokine release by MAPK-dependent histone modification. *J. Immunol.* 175, 2843–2850
- 33 Tsaprouni, L.G. *et al.* (2007) Suppression of lipopolysaccharide- and tumour necrosis factor-alpha-induced interleukin (IL)-8 expression by glucocorticoids involves changes in IL-8 promoter acetylation. *Clin. Exp. Immunol.* 150, 151–157
- 34 Gilmour, P.S. *et al.* (2003) Histone acetylation regulates epithelial IL-8 release mediated by oxidative stress from environmental particles. *Am. J. Physiol. Lung Cell Mol. Physiol.* 284, 533–540
- 35 Su, R.C. *et al.* (2009) Altered epigenetic regulation and increasing severity of bronchial hyperresponsiveness in atopic asthmatic children. *J. Allergy Clin. Immunol.* 124, 1116–1118
- 36 Ito, K. *et al.* (2002) Expression and activity of histone deacetylases in human asthmatic airways. *Am. J. Respir. Crit. Care Med.* 166, 392–396
- 37 Cosio, B.G. *et al.* (2004) Histone acetylase and deacetylase activity in alveolar macrophages and blood monocytes in asthma. *Am. J. Respir. Crit. Care Med.* 170, 141–147
- 38 Ito, K. *et al.* (2005) Decreased histone deacetylase activity in chronic obstructive pulmonary disease. *N. Engl. J. Med.* 352, 1967–1976
- 39 Adcock, I.M. *et al.* (2006) Abnormal histone acetylase and deacetylase expression and function in lung inflammation. *Inflamm. Res.* 55, 311–321
- 40 Mroz, R.M. *et al.* (2007) Molecular basis of chronic inflammation in lung diseases: new therapeutic approach. *J. Physiol. Pharmacol.* 58, 453–460
- 41 Rajendrasozhan, S. *et al.* (2009) Current perspectives on role of chromatin modifications and deacetylases in lung inflammation in COPD. *Copd* 6, 291–297
- 42 Miao, F. (2004) *In vivo* chromatin remodeling events leading to inflammatory gene transcription under diabetic conditions. *J. Biol. Chem.* 279, 18091–18097
- 43 Chen, S. *et al.* (2010) Transcriptional coactivator p300 regulates glucose-induced gene expression in endothelial cells. *Am. J. Physiol. Endocrinol. Metab.* 298, E127–137
- 44 Liu, B. and Hong, J.S. (2003) Role of microglia in inflammation-mediated neurodegenerative diseases: mechanisms and strategies for therapeutic intervention. *J. Pharmacol. Exp. Ther.* 304, 1–7
- 45 Zhu, W. *et al.* (2010) Excitotoxicity of TNFalpha derived from KA activated microglia on hippocampal neurons *in vitro* and *in vivo*. *J. Neurochem.* 114, 386–396
- 46 Qin, H. *et al.* (2005) LPS induces CD40 gene expression through the activation of NF-kappaB and STAT-1alpha in macrophages and microglia. *Blood* 106, 3114–3122
- 47 Ma, X. *et al.* (2010) IL-17 enhancement of the IL-6 signaling cascade in astrocytes. *J. Immunol.* 184, 4898–4906
- 48 Mattson, M.P. and Camandola, S. (2001) NF-kappaB in neuronal plasticity and neurodegenerative disorders. *J. Clin. Invest.* 107, 247–254
- 49 Wang, S.X. *et al.* (2010) Anti-inflammatory activity of salvianolic acid B in microglia contributes to its neuroprotective effect. *Neurochem. Res.* 35, 1029–1037
- 50 Hwang, J. *et al.* (2008) Anti-inflammatory effects of m-chlorophenylpiperazine in brain glia cells. *Int. Immunopharmacol.* 8, 1686–1694
- 51 Dekker, F.J. and Haisma, H.J. (2009) Histone acetyl transferases as emerging drug targets. *Drug Discov. Today* 14, 942–948
- 52 Rekowski, M.W. and Giannis, A. (2010) Histone acetylation modulation by small molecules: a chemical approach. *Biochim. Biophys. Acta* 1799, 760–767
- 53 Lau, D.O. *et al.* (2000) HATs off: selective synthetic inhibitors of Histone Acetyltransferase p300 and PCAF. *Mol. Cell* 5, 589–595
- 54 Wu, J. *et al.* (2009) Bisubstrate Inhibitors of the MYST HATs Esa1 and Tip60. *Bioorg. Med. Chem.* 17, 1381–1386
- 55 Stimson, L. *et al.* (2005) Isothiazolones as inhibitors of PCAF and p300 histone acetyltransferase activity. *Mol. Cancer Ther.* 4, 1521–1532
- 56 Dekker, F.J. *et al.* (2009) Inhibition of PCAF histone acetyl transferase and cell proliferation by isothiazolones. *Bioorg. Med. Chem.* 17, 460–466
- 57 Ghizzoni, M. *et al.* (2009) Reactivity of isothiazolones and isothiazolone-1-oxides in the inhibition of the PCAF histone acetyltransferase. *Eur. J. Med. Chem.* 44, 4855–4861
- 58 Gorsuch, S. *et al.* (2009) Synthesis of isothiazol-3-one derivatives as inhibitors of histone acetyltransferase (HATs). *Bioorg. Med. Chem.* 17, 467–474
- 59 Wisastra, R. *et al.* (2011) Isothiazolones; thiol-reactive inhibitors of cysteine protease cathepsin B and histone acetyltransferase PCAF. *Org. Biomol. Chem.* 9, 1817–1822
- 60 Bowers, E.M. *et al.* (2010) Virtual ligand screening of the p300/CBP histone acetyltransferase: identification of a selective small molecule inhibitor. *Chem. Biol.* 17, 471–482
- 61 Mai, A. *et al.* (2006) Small-molecule inhibitors of histone acetyltransferase activity: identification and biological properties. *J. Med. Chem.* 49, 6897–6907
- 62 Chimenti, F. *et al.* (2009) A novel histone acetyltransferase inhibitor modulating Gcn5 network: cyclopentylidene-(4-(4'-chlorophenyl)thiazol-2-yl)hydrazon. *J. Med. Chem.* 52, 530–536
- 63 Mai, A. (2009) Identification of 4-hydroxyquinolines inhibitors of p300/CBP histone acetyltransferases. *Bioorg. Med. Chem. Lett.* 19, 1132–1135
- 64 Balasubramanyam, K. *et al.* (2003) Small molecule modulators of histone acetyltransferase p300. *J. Biol. Chem.* 278, 19134–19140
- 65 Sung, B. *et al.* (2008) Anacardic acid (6-nonadecyl salicylic acid), an inhibitor of histone acetyltransferase, suppresses expression of nuclear factor-kappaB-regulated gene products involved in cell survival, proliferation, invasion, and inflammation through inhibition of the inhibitory subunit of nuclear factor-kappaBalpha kinase, leading to potentiation of apoptosis. *Blood* 111, 4880–4891
- 66 Souto, J.A. *et al.* (2008) Synthesis of benzamides related to anacardic acid and their histone acetyltransferase (HAT) inhibitory activities. *ChemMedChem* 3, 1435–1442
- 67 Ghizzoni, M. *et al.* (2010) Improved inhibition of the histone acetyltransferase PCAF by an anacardic acid derivative. *Bioorg. Med. Chem.* 18, 5826–5834
- 68 Eliseeva, E.D. *et al.* (2007) Characterization of novel inhibitors of histone acetyltransferases. *Mol. Cancer Ther.* 6, 2391–2398
- 69 Ravindra, K.C. *et al.* (2009) Inhibition of lysine acetyltransferase KAT3B/p300 activity by a naturally occurring hydroxynaphthoquinone, plumbagin. *J. Biol. Chem.* 284, 24453–24464
- 70 Sandur, S.K. (2006) Plumbagin (5-hydroxy-2-methyl-1,4-naphthoquinone) suppresses NF-kappaB activation and NF-kappaB-regulated gene products through modulation of p65 and Ikbalpha kinase activation, leading to potentiation of apoptosis induced by cytokine and chemotherapeutic agents. *J. Biol. Chem.* 281, 17023–17033
- 71 Balasubramanyam, K. *et al.* (2004) Curcumin, a novel p300/CBP-binding protein-specific inhibitor of acetyltransferase, represses the acetylation of histone/nonhistone proteins and histone acetyltransferase-dependent chromatin transcription. *J. Biol. Chem.* 279, 51163–51171
- 72 Arif, M. *et al.* (2010) Nitric oxide-mediated histone hyperacetylation in oral cancer: target for a water-soluble HAT inhibitor, CTK7A. *Chem. Biol.* 17, 903–913
- 73 Lin, Y.G. *et al.* (2007) Curcumin inhibits tumor growth and angiogenesis in ovarian carcinoma by targeting the nuclear factor-kappaB pathway. *Clin. Cancer Res.* 13, 3423–3430
- 74 Choi, K.C. *et al.* (2009) Epigallocatechin-3-gallate, a histone acetyltransferase inhibitor, inhibits EBV-induced B lymphocyte transformation via suppression of RelA acetylation. *Cancer Res.* 69, 583–592
- 75 Choi, K. *et al.* (2009) Gallic acid suppresses lipopolysaccharide-induced nuclear factor-kappaB signaling by preventing RelA acetylation in A549 lung cancer cells. *Mol. Cancer Res.* 7, 2011–2021
- 76 Adachi, S. (2009) (–)-Epigallocatechin gallate downregulates EGF receptor via phosphorylation at Ser1046/1047 by p38 MAPK in colon cancer cells. *Carcinogenesis* 30, 1544–1552
- 77 Balasubramanyam, K. *et al.* (2004) Polyisoprenylated benzophenone, garcinol, a natural histone acetyltransferase inhibitor, represses chromatin transcription and alters global gene expression. *J. Biol. Chem.* 279, 33716–33726
- 78 Ahmad, A. *et al.* (2010) Apoptosis-inducing effect of garcinol is mediated by NF-kappaB signaling in breast cancer cells. *J. Cell. Biochem.* 109, 1134–1141
- 79 Mantelingu, K. *et al.* (2007) Specific inhibition of p300-HAT alters global gene expression and represses HIV replication. *Chem. Biol.* 14, 645–657